

ANI™ PARVOSTICK

ANI™ PARVOSTICK is a sensitive and rapid stick test for the detection of parvovirus from faecal samples
Cat. No. 1-001-200 / 1-001-205

TEST PRINCIPLE

ANI™ Parvostick is a rapid one step qualitative immunoassay based on the immunochromatographic sandwich principle. The method employs a unique combination of highly specific anti-parvovirus antibody-dye conjugate (colloidal gold) and anti-parvovirus solid phase antibody to selectively identify parvovirus antigen with a high degree of sensitivity.

TEST COMPONENTS

ANI™ Parvostick, cat.no 1-001-200:

- 10 Disposable test sticks
- 10 Test tubes
- 10 Sampling swabs
- 0,5 ml Positive Control
- 5,0 ml ANI™ Parvo Buffer (phosphate buffered saline)

Instructions for use

ANI™ Parvostick, cat.no 1-001-205:

- 5 Disposable test sticks
- 5 Test tubes
- 5 Sampling swabs
- 0,5 ml Positive Control
- 5,0 ml ANI™ Parvo Buffer (phosphate buffered saline)

Instructions for use

Materials needed but not provided with the kit: timer, specimen container

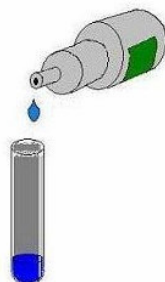
TEST PROCEDURE

1. Collection of the specimen

Faecal samples are collected using the cotton swabs from the anus, or e.g. from a thermometer after measuring temperature. Care should be taken to avoid heavy contamination of the swab sample with extraneous materials.

2. Handling of samples prior testing

Place a test tube in a workstation. Add 6 - 8 drops of ANI™ Parvo Buffer into the test tube.



Faecal specimens are collected using a cotton swab from the anus. The swab with a small amount of sample is put in the test tube. Please take care that not too much sample is caught on the inside of tube. The dilution should be appr. 1:10. The faecal sample is suspended in the buffer by rotating the swab.



Remove the swab from the tube. The test tube should stand at room temperature for 3 minutes to allow coarse particles to sediment. Samples can be stored for seven days refrigerated before the testing procedure.

3. Testing procedure

Take a stick from the aluminium foil pouch and place it standing in the test tube with the filter end in the sample. The stick should not be

submerged in the sample deeper than the length of the max line indicates.



Let the stick remain standing in the sample for 5 minutes.

4. Interpretation of the results

A positive result is indicated by the appearance of two red lines in the white central area of the stick. This can happen with clear positive samples in as soon as 2 minutes and with weak positive samples in 5-10 minutes.



A negative result is indicated by the appearance of one red line, the control line, only. This is also an indication of proper performance of the test.



If the control line is not visible the test is inconclusive, whether or not the test line is visible. A new test must be performed.

STORAGE

Test packages can be stored at ambient temperature (+2 ... +25 °C), until expiry date. ANI™ Parvo Positive Control should be kept refrigerated (+2 ... +8 °C).

CONTROLS

The proper performance of ANI™ Parvostick can be checked by means of ANI™ Parvo Positive Control or with a known faecal specimen. The positive control shall not be diluted further as it is ready for testing. The ANI™ Parvo Buffer used as a negative control shall produce a negative result. When known frozen specimens are used as controls, they should always be diluted with ANI™ Parvo Buffer.

SENSITIVITY AND SPECIFICITY

The sensitivity has been adjusted with standard faecal samples containing parvovirus antigen.

Ani Parvostick has no cross-reactivity with Adenovirus or Rotavirus.

If further information is needed, a documentation file is available.

WARNINGS AND LIMITATIONS

Follow the instructions for use carefully; a failure to do so may result in false test results. ANI™ Parvostick test may only be used for in-vitro qualitative detection of Parvovirus antigen from a faecal specimen according to the instructions for use.

Do not use the test if the foil pouch is damaged. Do not use expired tests or reagents. Only use the reagents that came with the kit. Use each test strip only once.

The test should be performed in accordance with generally accepted virological principles of handling and containment of potentially hazardous samples. All samples are potential pathogens. Used materials and samples should be disposed of with appropriate caution.

Sodium Azide has been added (0,09 %) to the reagents as a preservative. Avoid contact with skin and mucous membranes.

The positive control contains materials derived from animals. Handle with appropriate care.

REFERENCES

1. Neuvonen, E., Veijalainen, P.M.-L. and Kangas, J. 1982 Canine parvovirus infection in housed raccoon dogs and foxes in Finland. Vet. Rec. 8:448-449
2. Veijalainen, P.M.-L. Neuvonen, E. and Kangas, J. 1984 Parvovirus infection in blue foxes, p. 581-585 In communications, Proceedings. 3rd International Scientific congress in Fur Animal Production, Versailles.
3. Veijalainen, P.M.-L., Neuvonen, E., Niskanen, A. and Juokslahti, T. 1986 Latex agglutination test for detecting feline panleukopenia virus, canine virus and parvovirus of fur animals J.Clin.Microbiol. 23:556-559

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